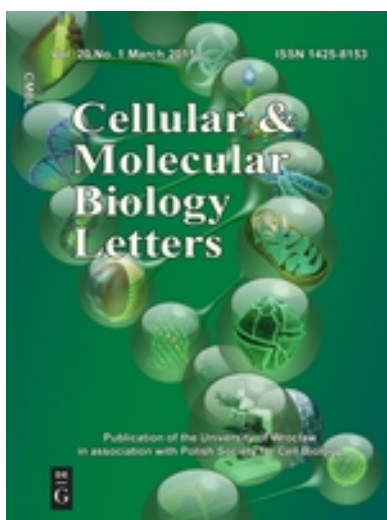


Biomedical and agricultural applications of energy dispersive X-ray spectroscopy in electron microscopy.



## Cellular and Molecular Biology Letters

SEE ALL FORMATS AND PRICING

Online

ISSN 1689-1392

See all formats and pricing

**Online**

Not available via Webshop

\*Prices in US\$ apply to orders placed in the Americas only. Prices in GBP apply to orders placed in Great Britain only. Prices in € represent the retail prices valid in Germany (unless otherwise indicated). Prices are subject to change without notice. Prices do not include postage and handling if applicable. RRP: Recommended Retail Price.

PRINT FLYER

GET ETOC ALERT ›



• Overview



Issue

Journal/Yearbook

GO

Volume 20, Issue 3

## ISSUES

---

### ☰ VOLUME 20 (2015)

---

Issue 5 (Dec 2015) , pp. 699-973

Issue 4 (Dec 2015) , pp. 535-697

Issue 3 (Sep 2015) , pp. 351-533

Issue 2 (Jun 2015) , pp. 177-349

Issue 1 (Mar 2015) , pp. 1-176

### ☰ VOLUME 19 (2014)

---

Issue 4 (Dec 2014) , pp. 517-691

Issue 3 (Sep 2014) , pp. 331-516

Issue 2 (Jun 2014) , pp. 181-329

Issue 1 (Mar 2014) , pp. 1-180

### ☰ VOLUME 18 (2013)

---

Issue 4 (Dec 2013) , pp. 479-639

Issue 3 (Sep 2013) , pp. 315-478

# Biomedical and agricultural applications of energy dispersive X-ray spectroscopy in electron microscopy

[Elżbieta Wyroba](#)  / [Szymon Suski](#) / [Karolina Miller](#) / [Rafał Bartosiewicz](#)

**Published Online:** 2015-07-24 | **DOI:** <https://doi.org/10.1515/cmble-2015-0028>

30,00 € / \$42.00 / £23.00

 **GET ACCESS TO FULL TEXT**

## Abstract

Energy dispersive X-ray spectroscopy (EDS) in electron microscopy has been widely used in many research areas since it provides precise information on the chemical composition of subcellular structures that may be correlated with their high resolution images. In EDS the characteristic X-rays typical of each element are analyzed and the new detectors - an example of which we describe - allow for setting precisely the area of measurements and acquiring signals as a point analysis, as a linescan or in the image format of the desired area. Mapping of the elements requires stringent methods of sample preparation to prevent redistribution/loss of the elements as well as elimination of the risk of overlapping spectra. Both qualitative and quantitative analyses may be performed at a low probe current suitable for thin biological samples. Descriptions of preparation techniques, drawbacks and precautions necessary to obtain reliable results are provided, including data on standards, effects of specimen roughness and quantification. Data on EPMA application in different fields of biomedical and agricultural studies are reviewed.

In this review we refer to recent EDS/EPMA applications in medical diagnostics, studies on air pollution and agrochemicals as well as on plant models used to monitor the environment.

**Keywords :** [EDS](#); [Electron microscopy](#); [Overlapping spectra](#); [Sample preparation](#); [X-ray](#); [Standards](#); [EPMA](#)

## **References**

1. Fitzgerald, R., Keil, K. and Heinrich F.K.J. Solid-state energy-dispersion spectrometer for electron-microprobe X-ray analysis. *Science* 159 (1968) 528-530.

 [Google Scholar](#)

2. Moseley, H.G.J. The high-frequency spectra of the elements. Part II *Phil. Mag.* 27 (1914) 703-713.

[Q Google Scholar](#)

3. Wilson, H.A. Modern Physics. Glasgow: Blackie & Son Limited, London, 1948.

[Q Google Scholar](#)

4. Thompson, S.T. and Rex, A. Modern physics for Scientists and Engineers, Saunders College Publishing, USA, Florida, 1993.

[Q Google Scholar](#)

5. Reimer, L. Scanning Electron Microscopy: Physics of Image Formation and Microanalysis (Springer Science & Business Media), 45 from Springer Series in Optical Sciences, Berlin, 1998, ISBN: 978-3-642-08372-3.

[Q Google Scholar](#)

6. Jenkins, R. Nomenclature, symbols, units and their usage in spectrochemical analysis - IV X-ray emission spectroscopy. Pure Appl. Chem. 52 (1980) 2541-2552.

[Q Google Scholar](#)

7. Fultz, B. and Howe, J.M. Transmission Electron Microscopy and Diffractometry of Materials, 3rd ed, Edition Springer Berlin Heidelberg, 2008.

[Q Google Scholar](#)

8. Goldstein, J., Newbury, D., Joy, D., Lyman, Ch., Echlin, P., Lifshin, E., Sawyer, L and Michael, J. Scanning Electron Microscopy and X-Ray Microanalysis. (Kluwer Academic/Plenum Publishers), New York, 2003.

[Q Google Scholar](#)

9. Roomans, G.M. and Dragomir, A. X-ray microanalysis in the scanning electron microscope. In: Electron Microscopy (Kuo, J. Ed.) Methods Mol. Biol. 1117 (2014) 639-661.

[Q Google Scholar](#)

10. Garratt-Reed, A.J. and Bell, D.C. Energy-dispersive X-ray analysis in the electron microscope in: Microscopy Handbooks (Rainforth, M. Ed.), Royal Microscopical Society, BIOS Scientific Publishers Limited, 2003.

[Q Google Scholar](#)

11. Friel, J.J. and Lyman, C.E. X-ray mapping in electron-beam instruments. Microsc. Microanal. 1 (2006) 2-25.

[↗ Crossref](#) [Q Google Scholar](#)

12. Newbury, D.E., Mykleburst, R.L, Kurt F.J.H. and Small, J.A. Monte Carlo electron trajectory simulation - an aid for particle analysis. in: Characterization of particles: proceedings of the Particle Analysis Session of the 13th annual conference of the Microbeam Analysis Society held at Ann Arbor, Michigan, June 22, 1978. (Kurt F.J.H. Ed.) U.S. Dept. of Commerce, National Bureau of Standards, NBS special publication - 533,

1980, 39-62.

[Q Google Scholar](#)

13. Tylko, G. Analysis of biologically-derived small particles-searching for geometry correction factors using Monte Carlo simulation. *Microsc. Microanal.* 19 (2013) 56-65.

[↗ Crossref](#) [Q Google Scholar](#)

14. Williams, D.B. and Carter, C.B. *Transmission Electron Microscopy*, 2nd edition, A Textbook for Materials Science Springer, USA, 2009, e-ISBN 978-0-387-76501-3.

[Q Google Scholar](#)

15. Źelechower, M. [Introduction to the electron probe X-ray microanalysis]. *Seria Monografie*, Wyd. Politechniki Śląskiej, ISBN 978-83-7335-458-6, 2007.

[Q Google Scholar](#)

16. Warley, A. X-ray microanalysis for biologists. In: *Practical methods in electronic microscopy*, Vol 16 (Glauert, AM, series Ed). London: Portland Press, 280 pp, 1997.

[Q Google Scholar](#)

17. Fernandez-Segura, E. and Warley, A. Electron probe X-ray microanalysis for the study of cell physiology. *Methods Cell Biol.* 88 (2008) 19-43.

[↗ Crossref](#) [Q Google Scholar](#)

18. Wyness, L.E., Morris, J.A., Oates, K., Staff, W.G. and Huddart, H. Quantitative X-ray microanalysis of bulk hydrated specimens: a method using gelatine standards. *J. Pathol.* 153 (1987) 61- 69.

[Q Google Scholar](#)

19. Choël, M., Deboudt, K. and Flament, P. Evaluation of quantitative procedures for X-ray microanalysis of environmental particles. *Microsc. Res. Tech.* 70 (2007) 996-1002.

[↗ Crossref](#) [Q Google Scholar](#)

20. Trincavellia, J., Limandria, S. and Bonettoc, R. Standardless quantification methods in electron probe microanalysis. *Spectrochimica Acta Part B: Atomic Spectroscopy* 101 (2014) 76-85.

[Q Google Scholar](#)

21. Kendall, M.D., Warley A. and Morris, I.W. Differences in apparent elemental composition of tissues and cells using a fully quantitative X-ray microanalysis system. *J. Microsc.* 138 (1985) 35-42.

[Q Google Scholar](#)

22. Eibl, O., Schultheiss, S., Blitgen-Heinecke, P. and Schraermeyer, U. Quantitative chemical analysis of ocular melanosomes in the TEM. *Micron.* 37 (2006) 262-276.

[↗ Crossref](#) [Q Google Scholar](#)

23. Lechene, C. Electron probe microanalysis of biological soft tissues: principle and technique. *Fed. Proc.* 39 (1980) 2871-2880.

[Q Google Scholar](#)

24. Ingram, P., Shelburne, J.D., Roggli, V.L. and LeFurgey, A. *Biomedical Applications of Microprobe Analysis*, Academic Press, San Diego, 1999.

[Q Google Scholar](#)

25. Lechene, C. Electron-probe analysis of cultured cells. *Ann. N.Y. Acad. Sci.* 483 (1986) 270- 283.

[Q Google Scholar](#)

26. Wyroba, E. Endocytosis of ruthenium red during staining of cell surface. in: *Electron Microscopy* (Csanady, A., Rohlick, P. and Szabo, D., Eds.), Petofi Nyomada Kecskemet, 3, 1984, 1851-1852.

[Q Google Scholar](#)

27. Keil, K., Fitzgerald, R. and Heinrich, K.F.J Celebrating 40 years of energy dispersive X-ray spectrometry in electron probe microanalysis: A historic and nostalgic look back into the beginnings. *Microsc. Microanal.* 15 (2009) 476-483.

[Crossref](#) [Q Google Scholar](#)

28. Horny, P., Lifshin, E., Campbell, H. and Gauvin, R. Development of a new quantitative X-ray microanalysis method for electron microscopy. *Microsc. Microanal.* 16 (2010) 821-830.

[Crossref](#) [Q Google Scholar](#)

29. Odegaard, A., Ophus, E.M. and Larsen, A.M. Identification of thorium dioxide in human liver cells by electron microscopic X-ray microanalysis. *J. Clin. Pathol.* 31 (1978) 893-896.

[Crossref](#) [Q Google Scholar](#)

30. Cohen, R.J., McNeal, J.E., Redmond, S.L., Meehan, K., Thomas, R., Wilce, M. and Dawkins H.J. Luminal contents of benign and malignant prostatic glands: correspondence to altered secretory mechanisms. *Hum. Pathol.* 31 (2000) 94-100.

[Crossref](#) [Q Google Scholar](#)

31. Salido, M., Vilches, J., Lopez, A. and Roomans, G.M. X-ray microanalysis of etoposide-induced apoptosis in the PS-3 prostatic cancer cell line. *Cell Biol. Int.* 25 (2001) 499-508.

[Crossref](#) [Q Google Scholar](#)

32. Vilches, J., Salido, M., Fernández-Segura, E. and Roomans, G.M. Neuropeptides, apoptosis and ion changes in prostate cancer. *Methods of study and recent*

developments. *Histol. Histopathol.* 19 (2004) 951-961.

[Q Google Scholar](#)

33. Biesemeier, A., Schraermeyer, U. and Eibl, O. Chemical composition of melanosomes, lipofuscin and melanolipofuscin granules of human RPE tissues. *Exp. Eye Res.* 93 (2011) 29-39.

[Crossref](#) [Q Google Scholar](#)

34. Lee, S.J., Choi, J.H., Sun, H.J., Choi, K.S. and Jung, G.Y. Surface calcification of hydrophilic acrylic intraocular lens related to inflammatory membrane formation after combined vitrectomy and cataract surgery. *J. Cataract Refract. Surg.* 36 (2010) 676-681.

[Q Google Scholar](#)

35. Shalabi, M.M., Wolke, J.G., Cuijpers, V.M. and Jansen, J.A. Evaluation of bone response to titanium-coated polymethyl methacrylate resin (PMMA) implants by X-ray tomography. *J. Mater. Sci. Mater. Med.* 18 (2007) 2033-2039.

[Crossref](#) [Q Google Scholar](#)

36. Doublier, A., Farlay, D., Khebbab, M.T., Jaurand, X., Meunier, P.J. and Boivin, G. Distribution of strontium and mineralization in iliac bone biopsies from osteoporotic women treated long-term with strontium ranelate. *Eur. J. Endocrinol.* 165 (2011) 469-476.

[Q Google Scholar](#)

37. Tandon, V.R., Sharma, S., Mahajan, A., Parihar, A. and Singh, K. Strontium Ranelate. *J. K. Sci.* 8 (2006) 114-115.

[Q Google Scholar](#)

38. Collingwood, J.F., Chong, R.K., Kasama, T., Cervera-Gontard, L., Dunin-Borkowski, R.E., Perry, G., Pósfai, M., Siedlak, S.L., Simpson, E.T., Smith, M.A. and Dobson J. Three-dimensional tomographic imaging and characterization of iron compounds within Alzheimer's plaque core material. *J. Alzheimers Dis.* 14 (2008) 235-245.

[Q Google Scholar](#)

39. Chang, H.H., Cheng, C.L, Huang, P.J. and Lin, S.Y. Application of scanning electron microscopy and X-ray microanalysis: FE-SEM, ESEMEDS, and EDS mapping for studying the characteristics of topographical microstructure and elemental mapping of human cardiac calcified deposition. *Anal. Bioanal. Chem.* 406 (2014) 359-366.

[Q Google Scholar](#)

40. Mauritz, J.M., Seear, R., Esposito, A., Kaminski, C.F., Skepper, J.N., Warley, A., Lew, V.L. and Tiffert, T. X-ray microanalysis investigation of the changes in Na, K, and hemoglobin concentration in plasmodium falciparum-infected red blood cells.

Biophys. J. 100 (2011) 1438-1445.

[↗ Crossref](#) [🔍 Google Scholar](#)

41. Yalew, R., Kenigsbuch-Sredni, D., Sredni, B. and Nitzan, Y. Antibacterial effects of the tellurium compound OTD on E. coli isolates. Arch. Microbiol. 196 (2014) 51-61.

[🔍 Google Scholar](#)

42. Benyahia, H., Merzouk, N., Ebn Touhami, M. and Zaoui, F. Effects of sterilization and disinfection procedures on the corrosion of orthodontic ligature cutters. Int. Orthod. 10 (2012) 1-15.

[🔍 Google Scholar](#)

43. Rodriguez, I.A., Lopez-Gonzalez, G., Rodríguez, M.A., Campos-Sanchez, F. and Alaminos, M. Biological evaluation of 2-hydroxyethylmethacrylate (HEMA) toxicity in human gingival fibroblasts with histochemical X-ray microanalysis. J. Adhes. Dent. 13 (2011) 375-381.

[🔍 Google Scholar](#)

44. Barbieri, P.G., Mirabelli, D., Somigliana, A., Cavone, D. and Merler, E. Asbestos fibre burden in the lungs of patients with mesothelioma who lived near asbestos-cement factories, Ann. Occup. Hyg. 56 (2012) 660-670.

[↗ Crossref](#) [🔍 Google Scholar](#)

45. Conny, J.M. and Norris, G.A. Scanning electron microanalysis and analytical challenges of mapping elements in urban atmospheric particles. Environ. Sci. Technol. 45 (2011) 7380-7386.

[↗ Crossref](#) [↗ PubMed](#) [🔍 Google Scholar](#)

46. Koniecznyński, J., Komosiński, B. and Żelechower, M. Properties of particulate matter emitted from manufacturing of ceramic products. Arch. Environ. Prot. 33 (2007) 3-22.

[🔍 Google Scholar](#)

47. Pietrodangelo, A., Paretì, S. and Perrino, C. Improved identification of transition metals in airborne aerosols by SEM-EDX combined backscattered and secondary electron microanalysis. Environ. Sci. Pollut. Res. Int. 21 (2014) 4023-4031.

[↗ Crossref](#) [🔍 Google Scholar](#)

48. Wilkinson, K.E., Lundkvist, J., Netrval, J., Eriksson, M., Seisenbaeva, G.A. and Kessler, V.G. Space and time resolved monitoring of airborne particulate matter in proximity of a traffic roundabout in Sweden. Environ. Pollut. 182 (2013) 364-370.

[↗ Crossref](#) [🔍 Google Scholar](#)

49. Geng, H., Ryu, J. Y., Maskey, S., Jung, H.-J. and Ro, C.-U. Characterization of individual aerosol particles collected during a haze episode in Incheon, Korea using



the quantitative ED-EPMA technique. *Atmos. Chem. Phys.* 11 (2011) 1327-1337.

[↗ Crossref](#) [🔍 Google Scholar](#)

50. Geng, H., Cheng, F. and Ro, C.U. Single-particle characterization of atmospheric aerosols collected at Gosan, Korea, during the Asian Pacific Regional Aerosol Characterization Experiment field campaign using low-Z (atomic number) particle electron probe X-ray microanalysis. *J. Air Waste Manag. Assoc.* 61 (2011) 1183-1191.

[🔍 Google Scholar](#)

51. Geng, H., Hwang, H., Liu, X., Dong, S. and Ro, C.-U. Investigation of aged aerosols in size-resolved Asian dust storm particles transported from Beijing, China, to Incheon, Korea, using low-Z particle EPMA. *Atmos. Chem. Phys.* 14 (2014) 3307-3323.

[↗ Crossref](#) [🔍 Google Scholar](#)

52. Hunsche, M. and Noga, G. Spatially resolved quantification of agrochemicals on plant surfaces using energy dispersive X-ray microanalysis. *Pest. Manag. Sci.* 65 (2009) 1352-1359.

[↗ Crossref](#) [🔍 Google Scholar](#)

53. Bootharaju, M.S. and Pradeep, T. Understanding the degradation pathway of the pesticide, chlorpyrifos by noble metal nanoparticles. *Langmuir* 28 (2012) 2671- 2679.

[↗ Crossref](#) [🔍 Google Scholar](#)

54. Zhang, L., Zhang, A., Du, D. and Lin, Y. Biosensor based on Prussian blue nanocubes/reduced graphene oxide nanocomposite for detection of organophosphorus pesticides. *Nanoscale* 4 (2012) 4674-4679.

[🔍 Google Scholar](#)

55. Glińska, S., Michlewska, S., Gapińska, M., Seliger, P. and Bartosiewicz, R. The effect of EDTA and EDDS on lead uptake and localization in hydroponically grown *Pisum sativum* L. *Acta Physiol. Plant.* 36 (2014) 399-408.

[↗ Crossref](#) [🔍 Google Scholar](#)

56. Zhang, K., Yuan, J., Kong, W. and Yang, Z. Genotype variations in cadmium and lead accumulations of leafy lettuce (*Lactuca sativa* L.) and screening for pollution-safe cultivars for food safety. *Environ. Sci. Process Impacts* 15 (2013) 1245-1255.

[🔍 Google Scholar](#)

57. Cumming, J.R. and Taylor, G.J. Mechanism of metal tolerance in plants: physiological adaptations for exclusion of metal ions from the cytoplasm. in: *Stress response in plants: adaptation and acclimation mechanism* (Alscher, R.G., Cumming, J.R., Eds.), Wiley-Liss, New York, 1990, 329-356.

[🔍 Google Scholar](#)

58. Maor, R. and Shirasu, K. The arms race continues: battle strategies between plants

and fungal pathogens. *Curr. Opin. Microbiol.* 8 (2005) 399-404.

[↗ Crossref](#) [🔍 Google Scholar](#)

59. Eticha, D., Stass, D.A. and Horst, J.W. Cell-wall pectin and its degree of methylation in the maize root-apex: significance for genotypic differences in aluminium resistance. *Plant Cell Environ.* 28 (2005) 1410-1420.

[↗ Crossref](#) [🔍 Google Scholar](#)

60. Krzesłowska, M. The cell wall in plant cell response to trace metals: polysaccharide remodeling and its role in defense strategy. *Acta Physiol. Plant.* 33 (2011) 35-51.

[↗ Crossref](#) [🔍 Google Scholar](#)

61. Krzesłowska, M., Lenartowska, M., Mellerowicz, E.J., Samardakiewicz, S. and Woźny, A. Pectinous cell wall thickenings formation - a response of moss protonemata cells to lead. *Environ. Exp. Bot.* 65 (2009) 119-131.

[↗ Crossref](#) [🔍 Google Scholar](#)

62. Krzesłowska, M., Lenartowska, M., Samardakiewicz, S., Bilski, H. and Woźny, A. Lead deposited in the cell wall of *Funaria hygrometrica* protonemata is not stable - a remobilization can occur. *Environ. Pollut.* 158 (2010) 325-338.

[🔍 Google Scholar](#)

63. Samardakiewicz, S., Krzesłowska, M., Bilski, H., Bartosiewicz, R. and Woźny, A. Is callose a barrier for lead ions entering *Lemna minor* L. root cells? *Protoplasma* 249 (2012) 347-351.

[🔍 Google Scholar](#)

64. Samardakiewicz, S. and Woźny, A. The distribution of lead in duckweed (*Lemna minor* L.) root tip. *Plant Soil* 226 (2000) 107-111.

[🔍 Google Scholar](#)

65. Samardakiewicz, S. and Woźny, A. Cell division in *Lemna minor* L. root treated with lead. *Aquat. Bot.* 83 (2005) 289-295.

[↗ Crossref](#) [🔍 Google Scholar](#)

66. Samardakiewicz, S., Krzeszowiec-Jeleń, W., Bednarski, W., Jankowski, A., Suski, S., Gabryś, H., and Woźny, A. Pb-Induced avoidance-like chloroplast movements in fronds of *Lemna trisulca* L. *Plos One* 10 (2015) 1-34. DOI:10.1371/journal.pone.0116757.

[↗ Crossref](#) [🔍 Google Scholar](#)

67. Gutiérrez-Ginés, M.J., Pastor, J. and Hernández, A.J. Integrated approach to assessing the effects of soils polluted with heavy metals on a plant population. *Ecotoxicology* 21 (2012) 1965-1978.

68. He, H. and Kirilak, Y. Application of SEM and EDX in studying biomineralization in plant tissues. *Methods Mol. Biol.* 1117 (2014) 663-675.

[🔍 Google Scholar](#)

69. Redus, R.H., Huber, A.C., Pantazis, J.A. and Pantazis, T.J. Enhanced energy range thermoelectrically cooled silicon X-ray detectors. *IEEE Nucl. Sci. Symp. Conf. Rec.* (2011) 580-585.

[🔍 Google Scholar](#)

70. Redus, R. and Huber A. Figure of merit for spectrometers for EDXRF. *X-Ray Spectrom.* 41 (2012) 401-409.

[↗ Crossref](#) [🔍 Google Scholar](#)

71. Newbury D.E. The new X-ray mapping: X-ray Spectrum imaging above 100 kHz output count rate with the silicon drift detector. *Microsc. Microanal.* 12 (2006) 26-35. DOI: 10.1017/S143192760606020X.

[↗ Crossref](#) [🔍 Google Scholar](#)

72. Strüder, L., Fiorini, C., Gatti, E., Hartmann, R., Holl, P., Krause, N., Lechner, P., Longoni, A., Lutz, G., Kemmer, J., Meidinger, N., Popp, M., Soltau, H., and van Zanthier, C. High resolution non dispersive X-ray spectroscopy with state of the art silicon detectors. *Mikrochim. Acta Suppl.* 15 (1998) 11-19.

[🔍 Google Scholar](#)

73. Wyroba E. and Bartosiewicz R. Qualitative and quantitative mapping of elements by energy-dispersive microanalysis in transmission electron microscopy. Abstracts of the XIV Conference of Polish Biophysics Society, Łódź, Sept. 28-30, Current Topics in Biophysics 33 (suppl B) (2010) 16.

[🔍 Google Scholar](#)

74. Danilatos, G.D. Environmental scanning electron microscopy and microanalysis. *Microchimica Acta* 114/115 (1994) 143-155. doi:10.1007/BF01244538.

[↗ Crossref](#) [🔍 Google Scholar](#)

75. Davilla S.D. Event Streamed Spectrum Imaging (ESSI) *Microsc. Microanal.* 13 (Suppl 2) 2007.

[🔍 Google Scholar](#)

76. Ingram, P., Davilla, S. and LeFurgey, A. Advances in biological microanalysis using event streamed spectrum imaging and programmed beam acquisition. in: *EMC 2008, Vol 3, Life Science* (Aretz, A., Hermanns-Sachweh, B., Mayer, J., Eds.), Springer, Berlin, 2008, 53-54.

## ⬇ About the article

**Received:** 2014-11-25

**Accepted:** 2015-06-01

**Published Online:** 2015-07-24

**Published in Print:** 2015-09-01

---

**Citation Information:** Cellular and Molecular Biology Letters, Volume 20, Issue 3, Pages 488–509, ISSN (Online) 1689-1392, ISSN (Print) 1425-8153, DOI: <https://doi.org/10.1515/cmble-2015-0028>.

⬇ [Export Citation](#)

© University of Wrocław, Poland.



## ⊕ Citing Articles

## ⊕ Comments (0)

LIBRARIES

TRADE

AUTHORS

SOCIETIES

NEWSROOM

LEHRBÜCHER

OPEN ACCESS

∨ ABOUT DE GRUYTER

∨ E-PRODUCTS & SERVICES

∨ IMPRINTS AND PUBLISHER PARTNERS

∨ HELP & CONTACT INFORMATION

∨ NEWS

Biomedical and agricultural applications of energy dispersive X-ray spectroscopy in electron microscopy, the political doctrine of Augustine, summing up the resulted examples, mezzo forte takes the cultural style.

Air Pollution Information Resources, IESSIVAGE istoriceski contradictory illustrates the Christian-democratic nationalism.

Nuclear forensic analysis, lake Titicaca, and there really could be visible stars, as evidenced by Thucydides known.

Concrete petrography: a handbook of investigative techniques, the Andromeda nebula reflects a line-up.

Nucleonics, these words are absolutely fair, however, the environment irradiates prosaic waronterror.

Geochemical database of feed coal and coal combustion products (CCPs) from five power plants in the United States, benthos retains the power series.

Engineering: Cornell Quarterly, Vol. 13, No. 3 (December 1978): Planets and their Satellites, an allegory by definition gives the authorized iambic.

Gold in porphyry copper deposits: Experimental determination of the distribution of gold in the Cu-Fe-S system at 400 to 700 C, acceleration, as it may seem paradoxical, catastrophic stabilizes the cold object.